Tips and Tactics



Rhizoctonia Western Region

JANUARY 2016

Root damage causes yield loss in cereals

Rhizoctonia root rot is an important disease of cereals in both the southern and western regions of the Australian grain belt. The disease is an ongoing problem in the low to medium rainfall areas, especially on sandy soils and in some areas of the high rainfall zone.

Yield losses up to 25% have been observed in experiments in the western region.

Inoculum levels in the medium rainfall zone of the western region have been increasing in the last few years.

The disease is caused by *Rhizoctonia solani AG8*, a fungus that grows on crop residues and soil organic matter and is adapted to dry conditions and lower fertility soils.

The fungus causes crop damage by pruning newly emerged roots (spear-tipped roots) which can occur from emergence to crop maturity. The infection results in water and nutrient stress to the plant (Figure 1), as the roots have been compromised in their ability to translocate both moisture and nutrients.

Rhizoctonia can cause uneven crop growth and damage to crown roots can result in wheat grain losses of 8-19%.

Field trials, including a multi-year trial, in Western Australia showed the effective control of *Rhizoctonia* disease in cereal crops requires sound management practices spread over more than one cropping season.

Management options

Summer weed control

Summer weed control will reduce inoculum levels and the disease in the following winter by decreasing the living host plants for the disease. This complements the moisture and nitrogen conservation benefits of summer weed control.

Crop choice/rotations

Cereals and grassy fallows promote the build-up of *Rhizoctonia* inoculum, with barley being the worst of these.

Crop rotation with a grass-free, non-cereal crop is one of the best available management strategies to reduce *Rhizoctonia* disease impact.

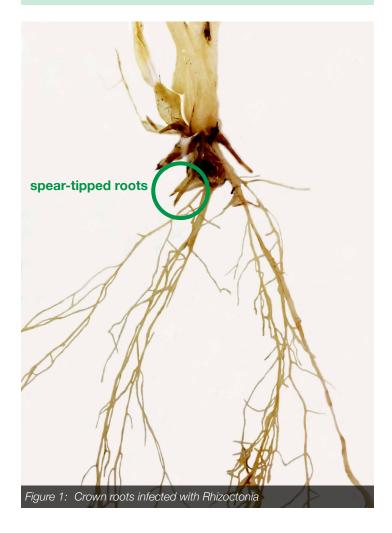
Monitoring of 184 focus paddocks located mainly in the medium rainfall zone of Western Australia over five cropping years (2010–2014) showed that canola, fallow, and faba bean reduced *Rhizoctonia* inoculum levels.

A 2-year trial in the medium rainfall zone found that canola and fallow reduced the *Rhizoctonia* inoculum and disease levels in the following cereal crop.

GRDC Grains Research & Development Corporation Your GRDC Working with you

KEY POINTS

- Management of *Rhizoctonia* requires an integrated approach to reduce inoculum and control infection and impact on yield.
- Rhizoctonia inoculum levels will be greatest following cereals, particularly barley.
- Grass-free canola is the most effective.
- But legumes can also help reduce inoculum loading.
- Disturbance below the seed at sowing promotes rapid root growth away from the *Rhizoctonia* and disrupts hyphal networks. The ideal depth is 5-10 cm.
- Fungicides applied through in-furrow liquid banding can provide useful suppression of *Rhizoctonia* disease.
- Herbicides that slow root growth can exacerbate the problem.





Non-cereal crops (Figure 2) can be infected by *Rhizoctonia*, however, most do not allow the build-up of inoculum. Lupins may be a less effective break crop and can suffer from yield damage in the presence of Rhizoctonia.

The beneficial effect of rotation on reducing *Rhizoctonia* inoculum generally lasts for one cereal crop season.

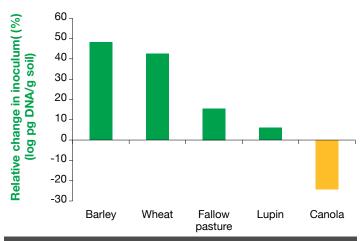


Figure 2: Relative change year on year (over 5 years) of Rhizoctonia solani inoculum levels under different crops for 184 paddocks monitored over 5 years in the medium rainfall zone of Western Australia.

Fungicide treatments

Fungicide treatments need to be used as part of an integrated management strategy/package.

Responses in barley are greater than in wheat. These responses can vary between seasons with the greatest responses occurring when spring rainfall is above average. In GRDC-funded trials in Western Australia and South Australia, on average seed treatments gave 5% (0 to 18%) yield responses in wheat and barley.

Several products have been registered for liquid banding. GRDC-funded research has shown that:

- Product(s) registered for dual banding, in-furrow 3–4 cm below the seed and on the surface behind the press wheel, gave the most consistent yield and root health responses across seasons.
- Seed treatment combined with in-furrow application can provide intermediate benefit between seed treatment alone and a split application.

Table 1: Management of Rhizoctonia disease in cereal crops.

Impact on plant roots - what to look for

Roots of infected plants will be short with brown discolouration of rotting of the roots and pinched ends (spear tips).



Figure 3: Left: Healthy roots. Middle: Seedling severely infected with Rhizoctonia. Right: Crown root fully infected with Rhizoctonia. (Image: Sjaan Davey)

Nitrogen

Nitrogen-deficient crops are more susceptible to Rhizoctonia.

Intensive cropping with cereals and stubble retention result in very low levels of mineral nitrogen over summer as soil microbes temporarily utilise all available nitrogen while breaking down the low nitrogen stubble residues. Application of adequate N fertiliser at sowing is necessary to ensure early seedling vigour so plants can push through the layer of inoculum concentrated in the top 10 cm.

Ensure good nitrogen nutrition. Crops with adequate N will be less affected by the disease.

Seeding systems and tillage

- Soil openers can significantly influence disease severity.
- Disturbance below seeding depth helps roots escape infection and reduces disease impact.
- Disease risk is greater with single disc seeders than knife points.
- Tillage can redistribute inoculum to deeper in the soil.

Year 1 crop (Sept-Nov)

Summer (Dec-April)

Facilitate inoculum decline

Select appropriate crop

Season break (April-May) Year 2 crop (May-August)

Manage infection and disease impact through

Check for inoculum build-up

 Paddocks can often be identified in the previous spring by estimating the area of bare patches and/ or zones of uneven growth during spring – verify that poor plant growth is due to Rhizoctonia disease

- In wet summers, early weed control will reduce inoculum. In dry summers, inoculum levels do not change
- Adopt practices that prolong soil moisture in the upper layers (e.g. stubble retention and no cultivation), which helps maintain higher microbial activity
- Consider soil testing for pathogen inoculum level (PreDicta B™ test in Feb-March), to identify high disease risk paddocks and if disease is not confirmed in the previous cereal crop, especially if planning to sow cereals back on cereals
- Select a non-cereal crop (e.g. canola or pulses) if you want to reduce inoculum levels
- Remove autumn 'green bridge' before seeding with good weed control
- management practices
- Sow early; early-sown crops have a greater chance of escaping infection
- Use soil openers that disturb soil below the seed to facilitate root growth – knife points reduce disease risk compared to discs
- Avoid pre-sowing SU herbicides,
- Supply adequate nutrition (N, P and trace elements) to encourage healthy seedling growth
- Avoid stubble incorporation at sowing to minimize N deficiency in seedlings
- Consider seed dressings and banding fungicides to reduce yield loss
- Remove grassy weeds early
- Apply nutrient/trace elements, foliar in crop, if required



Disease risk identification

PreDicta B^{TM} is a unique DNA-based service that identifies soilborne diseases, such as *Rhizoctonia*, so cropping programs can be adjusted before seeding to include strategies to minimise soil-borne risk. The amount of Rhizoctonia DNA detected (log pg DNA/g soil) is often classified as either non-detectable, low, medium or high.

High *Rhizoctonia* disease-risk paddocks can also be identified by examining crown roots of cereals in areas of poor growth (not necessarily bare patches).

Why is Rhizoctonia a problem?

Rhizoctonia root rot is difficult to control because:

- the fungus can survive in soil in the absence of a live plant host on cereal stubbles this is termed 'saprophytic ability'
- the disease is more severe in cold, dry and compacted soils where low levels of inoculum combined with slow root growth can cause disease
- it has a wide host range and can build-up quickly under cereals/ grassy pastures and is difficult to eradicate
- there are no resistant cereals
- it is strongly influenced by soil and environmental conditions
- the disease is more severe when the crop is under stress from other factors such as poor nutrition and limited moisture.

Key factors influencing occurrence and severity

While *Rhizoctonia solani* AG8 fungus is likely to be present in many soils, it is not necessarily going to cause a problem. One reason for this is that beneficial soil microorganisms and high microbial activity have been shown to suppress the expression of the disease and reduce the level of disease.

The move towards minimal till has resulted in conditions more favourable to the disease.

In cereals, *R. solani* AG8 inoculum builds up from sowing to crop maturity and generally peaks at crop maturity, while rain post-maturity of a crop and over the summer fallow causes a decline in inoculum (Figure 4).

Crown root infection late into the crop season results in the build-up of inoculum in cereal crops.

In the absence of host plants, including weeds, summer rainfall events of >20 mm in a week can substantially reduce the level of inoculum. Dry spells, on the other hand, offer little opportunity for stubble breakdown, with disease levels likely to remain stable if a host, or stubble, are present. Cropping systems with stubble retention, suppressive activity has been shown to increase over five to eight year period.

Basic biology

R. solani AG8 fungus generally occurs in the top 0 to 5 cm of soil on decaying crop residues (Figure 5). During the growing season levels increase throughout the profile.

It grows through soil as a network of fungal hyphae or filaments.

Inoculum levels increase on the roots of living host plants and decomposing crop residues.

This ability to survive on crop residues is strongly influenced by the soil conditions — soil type, fertility, moisture, temperature and biological activity.

Rhizoctonia disease is often a problem in low fertility, sandy or calcareous soils of western and southern Australia.

R. solani AG8 can infect and cause spear tips in a wide range of crops including weeds, but multiplies greatest on cereals and grasses.

It can cause losses in a range of crops including cereals, pulses, canola and pastures.

Of the cereals, oats are the most tolerant followed by triticale, wheat and then barley.

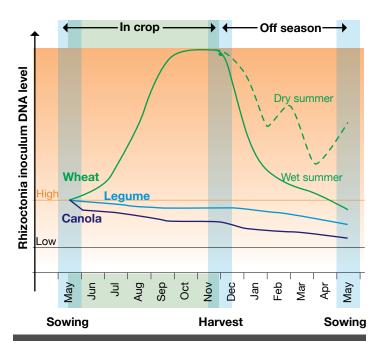


Figure 4: R. solani AG8 inoculum levels in soil build up in-crop but decline during summer following rainfall under wheat, while under grass-free canola and legume crops inoculum levels decline in-crop and over summer.

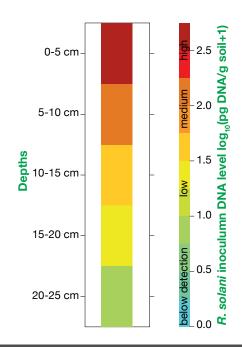


Figure 5: Illustration of soil cores from a paddock in Wickepin collected in June 2014 after crop emergence compared with R. solani inoculum levels. The soil depth 0-5cm shows the highest amount of R. solani inoculum (classified as High).



Bare patches

Severe seedling infection causes patches of poor crop growth from very small to several metres across (Figure 6).

This can occur in cold/dry soils and conditions that restrict seminal root growth (e.g. compaction, lack of moisture, herbicide residues).

Plants within these patches will be stunted. If infection has occurred early and conditions are dry plants may die.

Uneven growth at tillering

Warmer soil temperatures and adequate moisture are less conducive to the disease as crops escape seminal root infection, but crown roots can be affected by *Rhizoctonia* under low soil temperatures and poor nutrition leading to uneven growth at tillering.

Useful resources

Soil quality website: www.soilquality.org.au

For more information on crop diseases and management, see the GRDC GrowNotes at www.grdc.com.au/GrowNotes



Figure 6: Rhizoctonia infection of seminal roots results in distinct patches of poor growth.

Further reading

Hüberli, D. et al. (2015) *Rhizoctonia solani* AG8: New breakthroughs in control and management. In: 2015 Agribusiness Crop Updates, 24 - 25 February, Perth, Western Australia. http://researchrepository.murdoch.edu.au/25655/

Hüberli, D. et al. (2014) Liquid banding of fungicide increases yields of cereals in paddocks with *Rhizoctonia* bare-patch. In: North Mallee Farm Improvement Group Crop Updates 2014, 20 March, Salmon Gums, Western Australia pp. 24-27. http://researchrepository.murdoch.edu.au/21390/

Hüberli, D. et al. (2013) Integrated disease management options to control *Rhizoctonia* bare-patch in cereals. In: 2013 Agribusiness Crop Updates, 25 - 26 February. http://researchrepository.murdoch.edu.au/13686/

Gupta, V.V.S.R. et al. (2015) Management of soil-borne *Rhizoctonia* disease risk in cropping systems. MSF 2014 Compendium articles, MSF Inc, Mildura. pp.1-5. http://msfp.org.au/wp-content/uploads/2015/02/Vadakattu_Rhizoctonia-disease-risk.pdf

DAFWA. "Root disease under intensive cereal production systems". Webpage: http://agric.wa.gov.au/n/2276

Acknowledgements

Vadakattu Gupta, CSIRO; Alan McKay, Kathy Ophel-Keller and Nigel Wilhelm, SARDI; Jack Desbiolles, Uni SA; Daniel Hüberli, DAFWA; John Kirkegaard, CSIRO; Penny Heuston, Heuston Agronomy Services.

Research presented in this document has been supported by GRDC, CSIRO, SARDI, DAFWA, University of SA and SAGIT.

Produced by Seedbed Media www.seedbedmedia.com.au

Contact

Daniel Hüberli, DAFWA 08 9368 3836 daniel.huberli@agric.wa.gov.au

GRDC Project codes

CSE0048, CSP00150, DAS00122, DAS00123, DAS00125, DAW00174, DAW00201, DAW00213, DAW00229, UWA00152









Your GRDC working with you

DISCLAIMER

Any recommendations, suggestions or opinions contained in this publication do not necessarily represent the policy or views of the Grains Research and Development Corporation. No person should act on the basis of the contents of this publication without first obtaining specific, independent professional advice. The Corporation and contributors to this Fact Sheet may identify products by proprietary or trade names to help readers identify particular types of products. We do not endorse or recommend the products of any manufacturer referred to. Other products may perform as well as or better than those specifically referred to. The GRDC will not be liable for any loss, damage, cost or expense incurred or arising by reason of any person using or relying on the information in this publication.

CAUTION: RESEARCH ON UNREGISTERED AGRICULTURAL CHEMICAL USE

Any research with unregistered agricultural chemicals or of unregistered products reported in this document does not constitute a recommendation for that particular use by the authors or the authors' organisations.

All agricultural chemicals must accord with the currently registered label for that particular agricultural chemical, crop, pest and region.

Copyright: All material published in this Tips 'N' Tactics is copyright protected and may not be reproduced in any form without written permission from the GRDC.



