In-crop management – disease

Key points

- Start to monitor at early growth stages and continue to monitor for disease through the growing season
- Control foliar diseases when at early stages, preferably before canopy closure
- Use integrated approaches to pest, disease and weed management
7.1 Disease management

The diseases that cause major damage in vetch are chocolate spot (*Botrytis fabae*), Botrytis grey mould (*Botrytis cinerea*), rust (*Uromyces viciae-fabae*), Sclerotinia stem rot (*Sclerotinia sclerotiorum* and *S. trifoliorum*), stem nematode (*Ditylenchus dipsaci*), *Rhizoctonia* spp., and the mosaic and yellowing viruses. The other diseases listed in this section can cause problems in some seasons or regions (see Section 7.1.1 to Section 7.1.7).

Effective disease management relies on an integrated approach including: selection of a variety with the most suitable profile of disease resistance, most suitable paddock, clean seed, best agronomic practices and canopy management, as well as the use of fungicides (see Section 7, Table 1).

The use of integrated disease management (IDM) in vetch is especially important because controlling established foliar diseases with fungicides in vetch may not always be possible as there are few registered fungicides and some have long withholding periods (see Table 2). For rust there are several options, while control options for Ascochyta are limited and viruses non-existent.

If not sowing a resistant variety, a fungicide application may be needed in seed production crops, especially in high-rainfall regions, wet years or high disease risk situations (see Section 2.5.4).

Fungicides can be necessary to control rust in susceptible varieties if used for feed as infected plants can induce abortions in pregnant livestock.

Care needs to be taken using some fungicides that have long withholding periods (WHP) (for example, carbendazim, 28 days WHP) (see Table 2). Fungicides with long WHPs should be avoided if the vetch crop is to be conserved as silage or hay destined for the dairy industry. The withholding period must be completed before cutting because time elapsed after cutting does not count as being within the withholding period (http://agriculture.vic.gov.au/agriculture/farm-management/chemical-use/agricultural-chemical-use/chemical-labels/withholding-period-statements-on-labels).

Ascochyta blight occurs in the earlier stage of crop development, potentially reducing grain and dry matter production. However, Botrytis can cause greater yield losses if the crop is dense and the growing season is cool and wet (see Table 3). Note that some Ascochyta species are crop specific.1

Understanding the potential sources of disease inoculum is important when planning planting and control programs (see Section 2.1, Section 3.5 and Table 2).

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### Table 1: Integrated disease management strategies for vetch.

<table>
<thead>
<tr>
<th>Factor</th>
<th>Best practice</th>
</tr>
</thead>
<tbody>
<tr>
<td>Paddock history</td>
<td>A minimum of three, preferably four, years of break between vetch crops.</td>
</tr>
<tr>
<td>Paddock hygiene</td>
<td>Select paddocks at least 500 m from last year’s vetch stubble. Avoid sowing adjacent to faba bean and, if possible, field pea and lentil stubbles (see Section 2, Table 2, and Section 7, Table 2).</td>
</tr>
<tr>
<td>Variety</td>
<td>Select a variety with suitable disease resistance for your district. (see Section 2, Table 1).</td>
</tr>
<tr>
<td>Seed health</td>
<td>Use seed from crops that had a low disease severity, especially at podding. Laboratory seed tests can confirm disease levels. Use seed with less than 10% chocolate spot or 5% Ascochyta. Rust is not seed-borne (see Section 4.3 Seed quality).</td>
</tr>
<tr>
<td>Sowing time</td>
<td>Do not sow too early. Early emergence leads to excessive vegetative growth, early exposure to disease and early canopy closure, increasing foliar disease (see Section 4.1 Time of sowing).</td>
</tr>
<tr>
<td>Sowing rate</td>
<td>Higher than ideal seeding rates and plant populations can lead to a dense crop canopy and increased disease risk (see Section 4, Table 2).</td>
</tr>
<tr>
<td>Row spacing</td>
<td>Wider rows can delay canopy closure, reducing the risk of chocolate spot. Any increased lodging may increase the chance of foliar diseases (see Section 4.5.2 Row spacing).</td>
</tr>
<tr>
<td>Canopy</td>
<td>Delay sowing, reduce seeding rates or else graze or cut early-sown crops.</td>
</tr>
<tr>
<td>Fungicide application</td>
<td>Success depends on monitoring, correct disease identification, adequate coverage and timeliness of sprays with the correct fungicide. Seed: To reduce transmission of disease (helps control Ascochyta, Botrytis grey mould and seedling root rots). Foliar: Most effective when applied before or at first signs of disease and before rain. Protection lasts 10–12 days. Subsequent new growth is unprotected.</td>
</tr>
<tr>
<td>Aphid control</td>
<td>Early detection and control can reduce virus spread. Summer weed control, thicker crop density, stubble and minimal bare soil reduce the presence of aphids (see Section 6.1.2).</td>
</tr>
<tr>
<td>Harvest management</td>
<td>Early harvest reduces disease infection on the seed. Windrow or desiccate to enable earlier harvesting.</td>
</tr>
</tbody>
</table>

### Table 2: Fungicide active ingredients registered for use on vetch.

<table>
<thead>
<tr>
<th>Chemical</th>
<th>Withholding period</th>
<th>Ascochyta blight</th>
<th>Chocolate spot and Botrytis grey mould</th>
<th>Rust</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Grazing</td>
<td>Harvest</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mancozeb</td>
<td>14 days</td>
<td>28 days</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>Carbendazim</td>
<td>28 days</td>
<td>28 days</td>
<td>Not registered</td>
<td>✓</td>
</tr>
<tr>
<td>Metiram</td>
<td>21 days</td>
<td>42 days</td>
<td>✓</td>
<td>✓</td>
</tr>
</tbody>
</table>


### Table 3: Fungicide spraying program.

<table>
<thead>
<tr>
<th>Critical period</th>
<th>Disease</th>
<th>Fungicide</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>1st critical period</td>
<td>Ascochyta blight</td>
<td>Mancozeb or metiram</td>
<td>Early fungicide application can restrict early development and spread of disease. Use at first sign of infection. Use the higher rate for dense crops or if disease pressure is severe.</td>
</tr>
<tr>
<td>Early vegetative (6–8 weeks after emergence).</td>
<td>Rust</td>
<td>Mancozeb or metiram</td>
<td></td>
</tr>
<tr>
<td>Rust plus</td>
<td>Ascochyta blight</td>
<td>Mancozeb or metiram</td>
<td></td>
</tr>
<tr>
<td>2nd critical period</td>
<td>Ascochyta blight and/or rust plus</td>
<td>Chocolate spot</td>
<td>Mancozeb or metiram</td>
</tr>
<tr>
<td>Before canopy closure (13–16 weeks after emergence).</td>
<td>Chocolate spot</td>
<td>Carbendazim</td>
<td>If Ascochyta is detected, and/or chocolate spot appears in the crop canopy, and rain or high humidity are likely, apply fungicide if crop has sufficient yield potential.</td>
</tr>
<tr>
<td>Chocolate spot plus</td>
<td>Ascochyta blight</td>
<td>Carbendazim + Mancozeb or metiram</td>
<td>Apply a maximum of two consecutive applications at 14-day intervals. Apply a minimum of 100L/ha water.</td>
</tr>
<tr>
<td>3rd critical period</td>
<td>Ascochyta blight and/or rust plus</td>
<td>Chocolate spot</td>
<td>Mancozeb or metiram</td>
</tr>
<tr>
<td>End of flowering when pods are filling (15–20 weeks after emergence).</td>
<td>Chocolate spot plus</td>
<td>Ascochyta blight and/or rust</td>
<td>Carbendazim + Mancozeb or metiram</td>
</tr>
<tr>
<td>Chocolate spot</td>
<td>-</td>
<td>Carbendazim</td>
<td></td>
</tr>
</tbody>
</table>

7.1.1 Ascochyta blight (*Ascochyta fabae*)

**Description** – Dark leaf spots which show through both sides, becoming grey with age.

Leaf spots are circular, becoming elongated; pale centres may fall out, leaving holes in leaf; tiny black-fruiting bodies develop within lesions.

Herbicide damage (particularly simazine) can be mistaken for Ascochyta blight but is usually confined to leaf margins and spots do not have grey centres with black specks.

Stems develop elongated, dark, sunken lesions; stems may split and break causing plants to lodge.

Pods develop black, sunken lesions, which can penetrate the pod and infect the developing seed. Badly infected seeds have brown or black stains.

First appears on leaves of seedlings when wet, cold conditions occur, usually well before flowering. Progresses to infect upper leaves, flowers, stems and pods.

Infection on mature pods leads to seed staining, especially when late rains occur pre-harvest. Disease can develop on pods of windrowed crops.

This disease is widespread in southern Australia.

**Economic importance** – Mostly in high-rainfall areas, more than 450 mm per year, but severity varies greatly crop to crop and season to season. Yield losses are not as severe as Ascochyta blight in faba bean.²

**Management** – Grow varieties with resistance; sow disease-free seed; use crop rotation; sow away from vetch and bean residues (including self-sown plants).

Apply fungicide during seedling stages 6–8 weeks after sowing. Spray before an average infection of one lesion per plant develops and rainfall is likely during the next week. A late fungicide application after flowering and no new growth is occurring assists in preventing seed staining.

Sowing into standing stubble helps raise the crop off the ground and improve airflow, making the crop environment less conducive to Ascochyta.

7.1.2 Chocolate spot and Botrytis grey mould (*Botrytis fabae* and *B. cinerea*)

**Description** – These are major diseases in all vetch growing areas. They infect plants at any stage but are worse after flowering; they defoliate plants, ruining forage quality and reducing pod set.

**Botrytis** is favoured by temperatures of 15°C to 25°C and high humidity (>70%) for 4–5 days. Very rapid build-up (aggressive stage) during warm, humid conditions late in the season. Worse in early-sown and dense crops, and heavy wet soils. Fluffy, grey fungal growth produces masses of spores on fallen leaves and petals under wet conditions.

Non-aggressive spots, initially pinhead-sized and circular, reddish-brown on leaves and flowers; spots on one side of leaf only, most obvious as 'chocolate spots' early in the season. Spores are wind-blown so disease tends to be in the upper canopy. Spots expand rapidly and combine under suitable conditions, blackening and killing large areas of leaf; infection can spread into stems.

Flowers turn brown and are killed, reducing pod set. Pods develop reddish-brown, pinhead-sized spots. Pods may split, allowing infection of seed, which may be covered in small reddish-brown spots.

Severe infection can result in complete crop failure.

Economic importance – Occurs in all areas where vetch can grow. Losses range from minor to complete crop failure depending on the level of infection, time of infection occurrence and amount of spring rainfall. In unprotected crops, in a mild, wet spring disease commonly reduces yield by 30–50% through loss of leaves and collapse of stands.

Management – No varieties have good resistance. Fungus survives on stubble, volunteers and infected seed, so ensure good rotations and pre-seeding hygiene.

Manage the canopy by delaying sowing; using crop rotation and sowing away from bean and vetch residues. Control volunteer vetch plants. Check the crop every seven days when temperatures are below 15°C.

Check every three days when 15–20°C and humidity is over 70%.

Fungicide application may be futile if disease has developed unchecked and the crop is bulky and lodged, preventing fungicide penetration into the canopy.

Less than 350 mm/year
- graze crop to reduce canopy size if required, or
- apply a protective fungicide only if disease risk is high in a seed production crop and humidity in the crop is likely to be high for at least a week, especially if signs of infection are present.

More than 450 mm/year
- graze crop to reduce canopy size if required, or
- apply a protective fungicide if disease is present or risk is high in a grain production crop and humidity in the crop is likely to be high for at least a week.
- Repeat (10–21 days in severe cases) before rain, as determined by unprotected growth, rain since last application and expected rain. Last spray when flowering has ceased and no new growth is expected.

Photo 1: Dense, wet canopies, particularly in warm conditions, are conducive to the development of Botrytis, especially in the understorey where fungicides cannot penetrate.

Photo: Wayne Hawthorne, formerly Pulse Australia
7.1.3 Rust (Uromyces viciae-fabae)

Description – Rust is the most important disease of vetch in Australia. It is most prevalent in long-season districts and in warmer areas, such as northern NSW. It occasionally causes significant crop losses in southern areas in warm to hot (20–30°C), humid conditions.

Severe infection causes premature defoliation, resulting in reduced seed size.

Initial infection appears as creamy-coloured spore masses on leaves; replaced by orange-brown pustules surrounded by a light yellow halo; severely infected leaves wither and drop off.

Rust pustules on stems are similar but often larger than on leaves, and become darker as plants mature. Isolated rust pustules may develop on the pods.

Rust can occur earlier in the season than in other pulses but generally occurs during warm, humid conditions through grain filling. Can develop very quickly under favourable conditions. Above 20°C, rust generations are every 10 days.

Economic importance – This disease has caused losses of up to 30% on its own and in combination with chocolate spot can reduce yields by up to 50%.

Management – Sow a resistant variety. Sow away from vetch and bean residues and self-sown plants remaining after sowing.

Where broad-spectrum fungicides are used as treatments for other diseases, separate rust control is unlikely to be needed unless the product used does not control rust (such as carbendazim).

Graze the canopy to reduce crop bulk and its susceptibility to rust. Grazing rust-infected plants has caused abortion in pregnant stock.

Photo 2: Rust is the most serious disease of vetch in Australia and growing varieties with resistance is an important management strategy. Symptoms can occur early in the season as creamy-coloured spore masses on leaves which are replaced by orange-brown pustules surrounded by a light yellow halo as the disease progresses; severely infected leaves wither and drop off.

Photo: R Matic
7.1.4 Root rots (Fusarium, Phoma, Rhizoctonia and Pythium spp.)

**Description** – Plants are stunted and often die in patches. Leaves yellow and wilt before dying. Crown and stem bases are brown to black.

Roots are blackened and the root system is severely reduced. Lateral roots are short with tips rotted and, if infected with *Rhizoctonia*, these are often pointed.

**Management** – Control any green plant growth for at least several weeks prior to sowing; *Rhizoctonia* builds up on green plants. It is also more common in sandy infertile soils and in crops sown using minimal disturbance (such as disc seeders).

Shattering the soil below seed level at seeding will reduce *Rhizoctonia*.

Ensure adequate nitrogen and zinc nutrition.

Avoid situations where there is wet, cold weather with poor soil structure and free surface water. Also close rotations of vetch, faba bean or other pulses, especially field pea and chickpea, can increase the severity of *Rhizoctonia*.

7.1.5 Sclerotinia stem rot (*Sclerotinia sclerotiorum* and *S. trifoliorum*)

*Sclerotinia* has a very wide host range for species, including most broadleaf crops and weeds. Sclerotinia can build up in paddocks and the sclerotes can survive up to 10 years in soil, so will be a problem for other crops like canola, grown in the rotation.

**Description** – Affects isolated plants at any stage of growth. Plants wilt and collapse.

Infects stems, leaves or pods; young plants develop a slimy-wet rot at ground level.

Plants have a blackened base covered with fluffy, white fungal growth and are easily pulled from the soil.

Sclerotes (2–5 mm in diameter) form on the surface and in the centre of stems. Sclerotes are white at first, then turn black and hard.

Occurs where there is a high frequency of pulses and oilseeds in the crop sequence, high seeding rates or cool, wet conditions.

**Management** – Once established in a crop it is difficult to control. Lower seeding rates, wider row spacing and good weed control produce a more open crop that is less prone to disease.

Rotation with cereals will decrease soil inoculum levels.
Photo 3: Sclerotinia occurs where there is a high frequency of pulses and oilseeds in the crop sequence, high seeding rates, or cool, wet conditions. Rotations including cereals decrease inoculum levels and are an important part of an integrated disease management strategy, as in-crop control is difficult.

Photo: T Bretag

7.1.6 Stem nematode (Ditylenchus dipsaci)

Stem nematode is not a recognised problem in vetch. Vetch is a host but is relatively tolerant compared to pea, oats and canola.

Description – Patches of malformed and stunted plants. Leaves curled with water-soaked spots. Stems sometimes die back, turning reddish-brown from the base and stopping at a leaf.

Herbicide damage can produce similar symptoms.

Only occurs in parts of South Australia and Victoria and is worse in wet conditions. It has not been recorded in Western Australia.

Management – Sow nematode-free seed. Use Predicta® B DNA-based testing service to assess soil status before sowing. Do not introduce through infected straw. Avoid rotations of susceptible crops, such as oats, wild oats, pea, vetch, and some broadleaf weeds (such as bedstraw) that can increase nematode populations.

7.1.7 Viruses

Virus diseases are not regarded as significant in vetch. Some viruses are seed-borne, but most rely on living plant tissue to survive between seasons (green bridge). It is possible for vetch to be a host for viruses and to infect other crops, yet show no symptoms.

Bean yellow mosaic virus (BYMV), Clover yellow vein virus (CYVV), Pea seed-borne mosaic virus (PSbMV), Broad bean wilt virus (BBWV) are non-persistent, which means aphids lose their infectivity soon after feeding on healthy plants. So, aphids usually only spread them over short distances.

Bean leafroll virus (BLRV), Subterranean clover red leaf virus (SCRLV), Beet western yellows virus (BWYV), Subterranean clover stunt virus (SCSV) (yellowing or luteoviruses) are persistent, with the aphids remaining infected for life. These viruses can be spread by aphids over long distances. The relatively long feeding

time needed for the aphid to transmit the virus makes them responsive to control by insecticides.

Virus symptoms can include yellowing, leaf mottling or mosaics, stunting and tip distortion. Symptoms can easily be mistaken for herbicide damage, nutrient deficiencies, salinity effects or other abiotic factors. It is difficult to diagnose a virus just on field symptoms.

Growers are advised to seek expert advice. Crop patches or rings which increase over time often indicate the presence of a virus.

**IN FOCUS**

Presence of aphids may indicate symptoms are caused by a virus.

1. Seed-borne viruses can be controlled by sowing virus-free seed (less than 0.1% seed infection in high-risk areas, less than 0.5% seed infection in low risk areas). Infection can come from infected neighbouring crops.

2. Minimise aphid landing sites. Avoid bare soil as aphids land in crops where there is a clearly defined contrast in colour between bare soil and green foliage. Ensure good crop establishment, retain standing cereal stubble and produce a dense crop canopy.

3. Minimise herbicide stress, as stressed plants are more attractive to aphids.

4. Control in-crop weeds (potential sources of virus) and/or vectors early.